

References

1. Peker E, Kirimi E, Tuncer O, et al. Necrotizing fasciitis caused by *Staphylococcus epidermidis* in a neonate with extremely low birthweight. *J Dermatol* 2010;37(7):671-3.
2. Abbo O, Accadbled F, Guitard J, et al. Necrotizing fasciitis due to *Pseudomonas aeruginosa* in immunocompromised children. *Pediatr Blood Cancer* 2010;55(1):213-4.
3. Malbrain ML, Cheatham ML, Kirkpatrick A, et al. Results from the international conference of experts on intra-abdominal hypertension and abdominal compartment syndrome I: definitions. *Intensive Care Med* 2006;32(11):1722-32.
4. Cheatham ML, Malbrain ML, Kirkpatrick A, et al. Results from the international conference of experts on intra-abdominal hypertension and abdominal compartment syndrome II: recommendations. *Intensive Care Med* 2007;33(6): 951-62.
5. Baharestani MM. An overview of neonatal and pediatric wound care knowledge and considerations. *Ostomy Wound Manage* 2007; 53(6):34-6.
6. Aydin U, Ozgenel Y, Kanturk R. Vacuum-assisted closure therapy in newborn gangrene. *J Plast Reconstr Aesthet Surg* 2010;63(3):e277-9.

Cytokine Gene Polymorphisms in Childhood Dilated Cardiomyopathy: Interferon- gamma, Tumor Necrosis Factor-alpha and Transforming Growth Factor - beta 1 Genes Are Associated with the Disease in Turkish Patients

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Dilated cardiomyopathy (DCM) is a cardiac muscle disease with reduced left ventricular systolic function^[1]. Myocardial inflammation is the most common mechanism in the pathogenesis of cardiomyopathy in which cytokines may play an

important role^[2]. The objective of this study was to investigate the associations between tumor necrosis factor-alpha (TNF- α , -308), transforming growth factor-beta 1 (TGF- β 1, +10, +25), interleukin-10 (IL-10, -1082, -819, and -592), interleukin-6 (IL-6, -174), interferon-gamma (IFN- γ , +874) gene polymorphisms and DCM.

Sixteen children with DCM (3 months-13 years) and 21 healthy controls were tested for the cytokine genes with polymerase chain reaction-sequence-specific primers (PCR-SSP). In our results, TNF- α (-308) A allele was higher in DCM ($P=0.03$). The frequency of TNF- α (-308) GG genotype (low expression) was significantly decreased in DCM ($P=0.02$). The children with DCM had significantly higher frequencies of IFN- γ (+874) TT genotype (high expression) and allele T while TA genotype (intermediate expression) was lower in patients ($P=0.003$, $P=0.01$ and $P=0.04$, respectively). Haplotype analyses showed that TT/GG and TC/GG haplotypes of TGF- β 1 (high expression) were significantly decreased while TC/GC, CC/GG and TT/GC (intermediate expression) haplotypes were increased ($P=0.01$ and $P=0.04$, respectively). There was no association between IL-6 and IL-10 genotypes/haplotypes and DCM ($P>0.05$).

TNF- α is a strong proinflammatory and immunomodulatory cytokine that intervenes inflammatory diseases and is produced by activated macrophages^[3]. Frequency of TNF- α allele A was found high in DCM^[4]. TNF- α allele A (-308) was found over-expressed in patients with end-stage non-ischemic myocardial dysfunction^[5]. Tired et al did not find any association between TNF- α (-308) polymorphism and DCM^[6]. In our study, allele A of TNF- α (-308) gene was found susceptible to DCM, while GG genotype of TNF- α (-308) showed a protective effect against the disease.

The production or activities of several cytokines are modulated by IFN- γ ^[7]. The AA homozygosity of IFN- γ (+874) T/A polymorphism was associated with poor prognosis in idiopathic DCM in older patients^[2]. IFN- γ protected against the development of severe chronic myocarditis, pericarditis, and DCM after Coxsackievirus B3 infection by reducing mast cell degranulation, and the profibrotic cytokines (IL-4, IL-1 β , TGF- β 1) in the heart^[8]. In our study, the high expression of IFN- γ was found susceptible to DCM. We

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hypothesized that IFN- γ might play a possible role in the immuno-inflammatory process of childhood DCM, although it is not clear whether these act to preserve or protect against further inflammatory injury.

IL-6 is one of the proinflammatory cytokines with many systemic effects, including cardiovascular system^[9]. The GG and GC genotypes of IL-6 (-174) are associated with increased levels of IL-6, while CC with decreased expression^[10]. IL-6 levels were significantly associated with all outcomes of heart disease in adults^[11]. IL-6 (-174) polymorphism was associated with LVESD and LVEDD in DCM^[8]. Although allele C was higher in our patient group, there was borderline statistical significance between the groups ($P=0.0590$).

IL-10 is a regulatory cytokine which inhibits the production of IFN- γ and TNF- α and antagonizes the proinflammatory cytokine response^[12]. The diagnosis of DCM has been associated with a reduction in IL-10 plasma levels, indicating its protective role in cytokine activation^[13]. However, recent studies have suggested that IL-10 polymorphisms are not associated with DCM^[2,4], in agreement with our results.

TGF- β 1 is an anti-inflammatory cytokine that might play a major role in the immune modulation of heart function^[6]. TGF- β 1 expression is increased in the myocardium of patients with DCM^[14]. TGF- β 1 polymorphisms were correlated with better exercise capacity, and heart failure symptoms^[2]. Tired et al found no relationship between TGF- β 1 gene polymorphism and DCM^[6]. This study indicates that the high expression of TGF- β 1 had a protective effect against the DCM, while intermediate expression had susceptibility to the disease. Fairweather et al reported that IFN- γ protected against the development of DCM after infection by reducing profibrotic cytokines like TGF- β 1^[8].

We conclude that the increase in the expression of IFN- γ and TNF- α genes may be associated with the etiopathogenesis of DCM; however, the increase in the expression of TGF- β 1 gene may play a protective role against the development of this disease.

Key words: Cytokines; Dilated cardiomyopathy; Gene Polymorphism

References

1. Nugent AW, Daubeney DE, Chandras P, et al. The epidemiology of childhood cardiomyopathy in Australia. *N Engl J Med* 2003;348(17):1639-46.
2. Adamopoulos S, Kolokathis F, Gkouziouta A, et al. Cytokine gene polymorphisms are associated with markers of disease severity and prognosis in patients with idiopathic dilated cardiomyopathy. *Cytokine* 2011;54(1):68-73.
3. Vadlamani L, Iyengar S. Tumour necrosis factor α polymorphism in heart failure/cardiomyopathy. *Congest Heart Fail* 2004;10(6):289-92.
4. Ito M, Takahashi H, Fuse K, et al. Polymorphisms of tumor necrosis factor- α and interleukin 10 genes in Japanese patients with idiopathic dilated cardiomyopathy. *Jpn Heart J* 2000;41(2):83-91.
5. Densem CG, Hutchinson IV, Yonan N, et al. Tumour necrosis factor alpha gene polymorphism: a predisposing factor to non-ischaemic myocardial dysfunction? *Heart* 2002;87(2):153-5.
6. Tired L, Mallet C, Poirier O, et al. Lack of association between polymorphisms of eight candidate genes and idiopathic dilated cardiomyopathy. *J Am Coll Cardiol* 2000;35(1):29-35.
7. Schroder K, Hertzog PJ, Ravasi T, et al. Interferon-gamma: an overview of signals, mechanisms and functions. *J Leukoc Biol* 2004;75(2):163-89.
8. Fairweather D, Frisancho-Kiss S, Yujung SA, et al. Interferon- γ protects against chronic viral myocarditis by reducing mast cell degranulation, fibrosis, and the profibrotic cytokines transforming growth factor- β 1, interleukin-1 β , and interleukin-4 in the heart. *Am J Pathol* 2004;165(6):1883-94.
9. Fonseca JE, Santos MJ, Canhao H, et al. Interleukin-6 as a key player in systemic inflammation and joint destruction. *Autoimmun Rev* 2009;8(7):538-42.
10. Fishman D, Faulds G, Jeffery R, et al. The effect of novel polymorphism in the interleukin-6 (IL-6) gene on IL-6 transcription and plasma IL-6 levels, and an association with systemic onset juvenile chronic arthritis. *J Clin Invest* 1998;102(7):1369-76.
11. Cesari M, Penninx BW, Newman AB, et al. Inflammatory markers and onset of cardiovascular events: result from the health ABC study. *Circulation* 2003;108(19):2317-22.
12. Couper KN, Blount DG, Riley EM. IL-10: The master regulator of immunity to infection. *J Immunol* 2008;180(9):5771-7.
13. Adamopoulos S, Parissis JT, Paraskevaidis I, et al. Effects of growth hormone on circulating cytokine network, and left ventricular contractile performance and geometry in patients with idiopathic dilated cardiomyopathy. *Eur Heart J* 2003;24(24):2186-96.
14. Aharinejad S, Krenn K, Paulus P, et al. Differential role of TGF- β 1/bFGF and ET-1 in graft fibrosis in heart failure patients. *Am J Transplant* 2005;5(9):2185-92.